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Trichrome Masson Stain Kit

Product name: Trichrome Masson Stain Kit
Catalog No.: TASS01

Introduction:

Trichrome Masson Stain Kit is used for the detection of collagen fibers in tissues such as skin, heart, etc. on formalin-fixed, paraffin-embedded sections, and may be used for frozen sections as well. The collagen fibers will be stained blue and the nuclei will be stained black and the background is stained red.

Form:

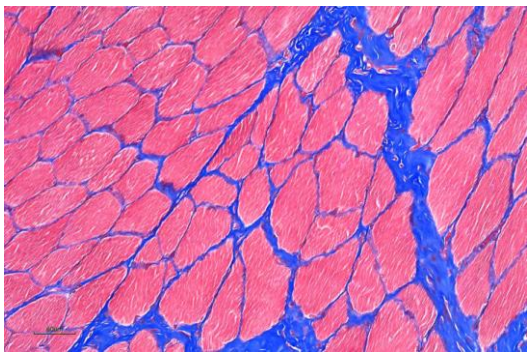
Catalog No.	Size
TASS01-125	125ml
TASS01-250	250ml

Kit Contents (for 250ml kit):

Kit Contents	Format	Recommend time	Storage
Biebrich scarlet-acid fuchsin solution	Ready to Use,250ml	10-15 minutes.	25-28℃
Phosphotungstic acid	Concentrate ,100ml	10-15 minutes.	25-28℃
Phosphomolybdic acid	Concentrate ,100ml	10-15 minutes.	25-28℃
Aniline blue solution	Ready to Use,250ml	3minutes.	25-28℃
Control slide x 2	Pig skin and others	--	25-28℃

Reagent necessary but not included:

1. 100% Alcohol
2. Bouin's solution
3. Weigert's iron hematoxylin.
4. 1% acetic acid solution
5. Acid alcohol.(4-5 drop 12N HCL in 180ml alcohol)



Preparing before use:

phosphomolybdic-phosphotungstic acid solution

(phosphomolybdic acid 1 part : phosphotungstic acid 1 part : dd Water 2 parts)

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Staining Protocol Recommendations:

1. Deparaffinize and rehydrate through 100% alcohol, 95% alcohol 70% alcohol.
2. Wash in distilled water.
3. For Formalin fixed tissue, re-fix in Bouin's solution for 1 hour at 60℃ or overnight at room temperature to improve staining quality although this step is not absolutely necessary.
4. Rinse running tap water for 5-10 minutes until the yellow color is removed.
5. Stain in Weigert's iron hematoxylin working solution for 5-10 minutes.
6. Wash in running tap water for 10 minutes and observe by eyes the tissue almost colorless. Rinse with distilled water. If necessary, you can use acidic alcohol to decolorize when background is blue or dark.
7. Stain in Biebrich scarlet-acid fuchsin solution for 10 minutes. If you want more vivid color, you can increase the staining time by 5 minutes. Solution can be reused for 2-3 times.
8. Flicking off the liquid from the slide.
9. Differentiate in phosphomolybdic-phosphotungstic acid solution for 3 minutes until collagen is clear (check the slide with microscopy).
10. Rinse briefly in distilled water.
11. Transfer the slide to aniline blue solution and stain for 3 minutes.
12. Rinse briefly in distilled water and differentiate in 1% acetic acid solution for 2 minutes.
13. Air dry or dehydrate very quickly through 95% ethyl alcohol, absolute ethyl alcohol (Alcohol will wipe off Biebrich scarlet-acid fuchsin staining) and clear in xylene.
14. Mount with resinous mounting medium.

Results:

Collagen ---------- blue
Nuclei ---------- black
Muscle, cytoplasm, keratin ---------- red

Positive Controls:

Skin, lung, stomach, intestine.

Storage and Stability:

Please read the kit contents and follow the storage condition. The user must validate any other storage conditions. When properly stored, the reagent is stable until the date indicated on the label. Do not use the reagent beyond the expiration date. If unexpected results are observed which cannot be explained by variations in laboratory procedures and a problem with the reagent is suspected, contact Technical: info@biotna.net